Falcon[®] Cell Culture Multi-Flask Guidelines for Use

INTRODUCTION

Falcon Cell Culture Multi-Flasks are available in 3- and 5-layer formats and provide 525 cm² or 875 cm² cell growth surface areas, respectively, which is equivalent to 3 or 5 times the surface area of a T-175 flask. Falcon Multi-Flasks offer the same benefits as the current format but in a more efficient and higher capacity design enabling you to grow more cells faster, easier, and in a more convenient manner. The footprint is the same as the T-175, so adaptation to your current equipment should be simple. This device allows you the benefit of preparing your cell suspension within the flask and removing your media or reagents using a pipet. This reduces the risk of contamination and speeds up the procedure.

ADDING MEDIA AND PREPARING CELL SUSPENSION WITHIN THE FALCON MULTI-FLASK



- **1.** Add required amount of medium into Falcon Multi-Flask by pipet or by pouring using typical culture volumes of 25-50 mL per layer.
- **2.** To avoid foaming of medium, allow liquid stream to flow along the slope of the Logo side of the Falcon Multi-Flask.

Helpful hint: A 10 mL pipet allows media to be dispensed at the bottom of the vessel. 25-100 mL pipets allow media to be dispensed just past the Logo.

MIXING OF CELLS IN FALCON MULTI-FLASK



4. Mix position: Hold the Falcon Multi-Flask upright with the Logo facing you and tilt counter-clockwise to a 45° angle.



- 5a. Holding at the same angle, gently rotate the Falcon Multi-Flask forward (neck pointing away from you).
- **5b.** Then, gently rotate it backward (neck pointing towards you).

Note: With each tilt, hold until liquid in the top layer drains fully.



 Repeat Step 5 to ensure proper mixing. Bring back to mix position, as shown in Step 4. Then, proceed to Step 7 for equilibration.



3. Dispense cell suspension from a concentrated stock into the growth medium using a Falcon 10 mL pipet (Cat. No. 357551 or 356551). Be sure to dip the pipet tip into the medium.

Note: The seeding density will vary depending on the cell type, medium, and culture duration needs. Begin with the same seeding density on a cells per centimeter square area to that used in standard flask for the cell type used.

EQUILIBRATING FLUID



7. After mixing or adding cell suspension, place the Falcon Multi-Flask vertically on a flat work surface to equalize liquid volume among all the layers.

ALTERNATE PROTOCOL

Adding cell suspension prepared external to the Falcon Multi-Flask

- 1. Create cell suspension externally from the Falcon Multi-Flask.
- Add required amount of cell suspension into the Falcon Multi-Flask by pipet or by pouring.
- To avoid foaming of medium, allow liquid stream to flow along the slope of the Logo side of the Falcon Multi-Flask.



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8. Hold the Falcon[®] Multi-Flask upright with the Logo facing you and tilt clockwise to a 45° angle on a flat work surface to partition the liquid into each of the layers. This position is recommended for transport.



9. While holding the Falcon Multi-Flask at a 45° angle, gently lay it flat onto the work surface with Logo facing up.



10. After placing the Falcon Multi-Flask flat on a work surface, gently rock back and forth and side-to-side to distribute cells evenly onto culture surfaces – taking care not to spill liquid from each layer.

PRODUCT SPECIFICATIONS

WORKING VOLUME RANGE	\geq 5 mL per layer for dissociating \leq 50 mL per layer for cell expansion
MOLDED-IN GRADUATIONS	0 to 50 mL per layer in 10 mL increments
GRADUATION ACCURACY	10%
CAP VENT MEMBRANE	0.2 µm hydrophobic membrane
CELL GROWTH SURFACE	Tissue culture-treated, optically clear

ORDER INFORMATION

Falcon Cell Culture Multi-Flask

Description	Qty./Pack	Qty./Case	Cat. No.
3-LAYER TISSUE CULTURE-TREATED, 525 CM ²	2	12	353143
5-LAYER TISSUE CULTURE-TREATED, 875 CM ²	1	8	353144

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To place an order in Europe, contact Customer Service at:

tel: + 31 0 20 659 60 51, fax: + 31 0 20 659 76 73; email: CSEurope@corning.com

For technical assistance, contact Technical Support at: email: **ScientificSupportEMEA@corning.com**

Outside Europe, contact your local distributor or visit www.corning.com/lifesciences to locate your nearest Corning office.

ACCESSORIES

Description	Qty./Pack	Qty./Case	Cat. No.
Falcon Pipets			
ASPIRATING PIPET 2 ML INDIVIDUALLY WRAPPED	50	200	357558
10 ML INDIVIDUALLY WRAPPED PAPER-PLASTIC	50	200	357551
10 ML INDIVIDUALLY WRAPPED ALL-PLASTIC	50	200	356551
Falcon Tubes			

50 ML POLYPROPYLENE CONICAL TUBE	25	500	352070
175 ML POLYPROPYLENE CONICAL TUBE	8	48	352076
225 ML POLYPROPYLENE CONICAL TUBE	8	48	352075

U.S. Orders

Contact your authorized distributor to place your order.

CORNING

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MEDIA REMOVAL

If exchange of media is required, follow Steps 7-10. You may choose to either aspirate or pour the media from the Falcon Multi-Flask.

ASPIRATING METHOD



11. To aspirate or remove media tilt the Falcon Multi-Flask, with the Logo facing you, counter-clockwise to a 45° angle while inverting the Multi-Flask toward you.

12. Then, tilt the Falcon Multi-Flask to the right, continuing to aspirate all residual media.

Helpful hint: Aspirate media using a Falcon 2 mL aspirating pipet (Cat. No. 357558).

POURING METHOD



13. With Logo facing down, pour spent media from the Falcon Multi-Flask.

Helpful hint: Pouring is easier when the Logo is facing you and the multi-flask is tilted at a counter-clockwise 45° angle.

CELL HARVESTING



- **14.** Add dissociating reagent (≥ 5 mL per layer) based on preferred protocol and bring to mix position (Step 4). Then, follow Steps 7-10.
- **15.** Neutralize with inactivating solution and mix following Steps 4-10. Gently swirl to dislodge cells completely.
- Pipetting Method: Follow "Media Removal" protocol but collect cell suspension using a Falcon 10 mL serological pipet (Cat. No. 357551).



- **17.** Follow Step 13 "Pouring Method". Pour detached cell suspension into a Falcon conical tube (Cat. No. 352070).
- **18.** Rinse with additional media as needed.

Helpful hint: Pouring is easier when the Logo is facing you and the multi-flask is tilted at a counter-clockwise 45° angle.

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