# Imaging of Cellular Fluorescence on Corning<sup>®</sup> Transwell<sup>®</sup> Permeable Supports

### **Application Note**

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### Introduction

Corning Transwell permeable supports can be used for a variety of cell culture applications to better mimic the in vivo environment. Transwell inserts with larger pore sizes, such as 3.0, 5.0, or 8.0  $\mu$ m, are typically used for migration, chemotaxis, and invasion assays, while Transwell inserts with smaller pore sizes, such as 0.4, 1.0, or 3.0 µm, can be used for co-culture applications and cell polarization studies. In co-culture and cell polarization applications, fluorescent imaging of cells cultured on the permeable membrane may be required to confirm cell membrane integrity or the presence of differentiated cell types. Traditionally, fluorescent imaging of cells cultured on Transwell inserts requires the membranes to be cut out of the Transwell insert and placed onto a slide or onto the bottom of a multiwell plate. However, with improvements in the capabilities and resolution of current imaging systems, especially confocal imagers, this may no longer be necessary. Here, we demonstrate fluorescent imaging of MDCKII/MDR1 cells directly on Transwell permeable supports in their cell culture receiver plate. This study utilized Transwell permeable supports with both polyester (PET) and polycarbonate (PC) membranes with pore sizes ranging from 0.4 to 3.0 µm.

#### **Materials and Methods**

MDCKII/MDR1 cells were obtained from Dr. Piet Borst (Netherlands Cancer Institute) and seeded into HTS 96-well Transwell permeable supports and 6.5 mm Transwell permeable supports at 1 x 10<sup>5</sup> cells/cm<sup>2</sup> in Dulbecco's Modified Eagle's Medium (DMEM; Corning 10-013-CM) supplemented with 10% Fetal Bovine Serum (FBS; Corning 35-010-CV). Transwell permeable support membranes tested included 0.4, 1.0, and 3.0  $\mu m$  PET as well as 0.4 and 3.0  $\mu m$ PC. The cells were cultured overnight in a 37°C, 5% CO<sub>2</sub> humidified incubator.

The next day, cells were stained live with 5  $\mu$ M Calcein AM (Corning 354216), 350 nM MitoTracker® Orange CMTMRos (Thermo Fisher M7510), and 10 µg/mL Hoechst 33342 (Thermo Fisher 62249) in Hank's Balanced Salt Solution with calcium and magnesium (HBSS; Corning 21-023-CM) for 45 minutes in a 37°C, 5% CO<sub>2</sub> humidified incubator. Cells were rinsed with HBSS prior to imaging with a Thermo Fisher CellInsight™ CX7 High-Content Screening (HCS) platform with a 20X objective in confocal mode, with 5 z-stack images of 2 µm steps taken for each representative composite image.

### **Results and Discussion**

Pore

Size

0.4 µm

1μm

3 µm

Calcein AM

Fluorescently labeled cells were imaged directly on each membrane type. Live cell stain Calcein AM (green), mitochondrial health stain MitoTracker Orange CMTMRos (red), and nuclear stain Hoechst 33342 (blue) were imaged in confocal mode and representative composite images are shown in Figure 1. The data demonstrates that removal of the PET and PC membranes from their supports is unnecessary for imaging, thus facilitating high throughput fluorescence microscopy of cells grown in Transwell permeable supports.

MitoTracker



cells stained live with Calcein AM (green), MitoTracker Orange CMTMROs (red), and Hoechst 33342 (blue). Cells were stained and imaged directly in Transwells on a variety of membrane types.

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#### **Recommendations for Protocol Optimization**

- Follow fluorescent staining protocols typically used for live or fixed cell staining.
- Create a new form factor in your imager specifically for Corning<sup>®</sup> Transwell<sup>®</sup> permeable supports. Start with a Corning 24-well or 96-well form factor and adjust the Z-height. The height may be outside of the range of auto-focus and need to be manually focused. To focus and find the correct Z-height for Transwell permeable supports, we recommend staining cells cultured on a Transwell permeable support with crystal violet or staining the cell nuclei.
- To accommodate for the increased distance between the cells and the imager, the exposure time may need to be increased compared with imaging of cells cultured on a microplate.

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